



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 6 : C12Q 1/68, C07H 21/00 // C12N 15/00, 14/00		A1	(11) International Publication Number: <b>WO 99/57309</b> (43) International Publication Date: 11 November 1999 (11.11.99)
(21) International Application Number:	PCT/DK99/00245	(81) Designated States:	AE, AL, AM, AT, AT (Utility model), AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, CZ (Utility model), DE, DE (Utility model), DK, DK (Utility model), EE, EE (Utility model), ES, FI, FI (Utility model), GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SK (Utility model), SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).
(22) International Filing Date:	4 May 1999 (04.05.99)	(30) Priority Data:	0615/98 4 May 1998 (04.05.98) DK
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(74) Agent: HOFMAN-BANG & BOUTARD, LEHMANN & REE A/S; Hans Bekkevolds Allé 7, DK-2900 Hellerup (DK).	(54) Title: METHOD AND PROBES FOR THE DETECTION OF CHROMOSOME ABERRATIONS		
(57) Abstract	<p>A novel method for detecting chromosome aberrations is disclosed. More specifically, chromosome aberrations are detected by <i>in situ</i> hybridisation using at least two sets of hybridisation probes, at least one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences related to a potential aberration in a chromosome, and at least one set comprising two or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences related to another potential aberration in a chromosome. In particular, the method may be used for detecting chromosome aberrations in the form of breakpoints.</p>		

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METHOD AND PROBES FOR THE DETECTION OF CHROMOSOME ABERRATIONS

The present invention relates to a novel method for 5 detection of chromosome aberrations. More specifically, the present invention relates to the determination of chromosome aberrations by *in situ* hybridisation using at least two sets of hybridisation probes, at least one set comprising one or more peptide nucleic acid probes 10 capable of hybridising to specific nucleic acid sequences related to a potential aberration in a chromosome, and at least another set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences related to another or the same potential 15 aberration in a chromosome. In particular, the present invention relates to the determination of chromosome aberrations by *in situ* hybridisation using at least two sets of hybridisation probes, at least one set comprising one or more peptide nucleic acid probes capable of 20 hybridising to specific nucleic acid sequences flanking one side of a potential breakpoint in a chromosome, and at least another set comprising two or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences flanking the other side of the 25 potential breakpoint in the chromosome. The invention further relates to such sets of hybridisation probes.

BACKGROUND OF THE INVENTION

30 Chromosome aberrations are well recognised to play an important role as a cause for diseases, not least as a cause for malignant diseases. Detection of such aberration is, therefore, an important target for novel diagnostic tools. Well known techniques known in the arts 35 of molecular biology and chromosome analyses are thus currently applied for detection of these important causes

of diseases. So far, however, a fast, convenient, cheap and user-friendly test which allows for widespread and routine detection of chromosome aberrations has not been available.

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According to i.a. the international patent application, publication no. WO 93/24652 (ref. 1) it has been found that peptide nucleic acids hybridise fast, selectively and strongly to nucleic acids such as, for example, 10 chromosomes or nucleic acids derived from chromosomes.

According to the international patent application, publication no. WO 97/18325 (ref. 2) i.a. chromosome aberrations can be detected by hybridising peptide 15 nucleic acids to chromosomes or fragments thereof followed by detection of the binding pattern of the peptide nucleic acids.

An area of particular interest regarding chromosome 20 aberration are the so called breakpoints that have been demonstrated in numerous cases to be specifically linked to the development of malignant diseases.

#### SUMMARY OF THE INVENTION

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In the broadest aspect, the present invention provides a method of determining the presence of chromosome aberrations in a sample of eukaryotic origin using *in situ* hybridisation, comprising the steps of 30

30

- a) producing a preparation of the sample, whereby the sample will be subject to a fixation,
- b) contacting said preparation with a hybridisation 35 solution comprising at least two sets of hybridisation probes, at least one set comprising one

or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences related to a potential aberration in a chromosome, and at least one set comprising one or more peptide 5 nucleic acid probes capable of hybridising to specific nucleic acid sequences related to another or the same potential aberration in a chromosome, and optionally a hybrid destabilising agent in an amount effective to decrease the melting temperature of 10 hybrids formed between said nucleic acid sequences and said peptide nucleic acid probes so as to increase the ratio between specific and non-specific binding,

15 c) removing any unbound and any non-specifically bound peptide nucleic acid probe, and  
d) determining the presence of peptide nucleic acid probe in the preparation.

20 In particular, the present invention enables the determination of chromosome aberrations in the form of breakpoints.

25 Thus, the present invention further provides a method of determining the presence of chromosome aberrations in a sample of eukaryotic origin using *in situ* hybridisation, comprising the steps of

30 a) producing a preparation of the sample, whereby the sample will be subject to a fixation,  
b) contacting said preparation with a hybridisation solution comprising at least two sets of 35 hybridisation probes one set comprising, at least one or more peptide nucleic acid probes capable of

hybridising to specific nucleic acid sequences flanking one side of a potential breakpoint in a chromosome, and at least one set comprising one or more peptide nucleic acid probes capable of

5 hybridising to specific nucleic acid sequences flanking the other side of the potential breakpoint and optionally a hybrid destabilising agent in an amount effective to decrease the melting temperature of hybrids formed between said nucleic acid sequences

10 and said peptide nucleic acid probes so as to increase the ratio between specific and non-specific binding,

c) removing any unbound and any non-specifically bound peptide nucleic acid probe, and

15 d) determining the presence of peptide nucleic acid probe in the preparation.

20 In a further aspect, the present invention provides a method of determining the presence of chromosome aberrations in a sample of eukaryotic origin using *in situ* hybridisation, comprising the steps of

25 a) producing a preparation of the sample, whereby the sample will be subject to a fixation,

b) contacting said preparation with a hybridisation solution comprising at least two sets of

30 hybridisation probes one set comprising, one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences flanking one side of a potential breakpoint in a chromosome, and one set comprising one or more peptide nucleic acid probes capable of hybridising to

35 specific nucleic acid sequences flanking the other

side of the potential breakpoint and optionally a hybrid destabilising agent in an amount effective to decrease the melting temperature of hybrids formed between said nucleic acid sequences and said peptide

5 nucleic acid probes so as to increase the ratio between specific and non-specific binding,

c) removing any unbound and any non-specifically bound peptide nucleic acid probe, and

10 d) determining the presence of peptide nucleic acid probe in the preparation.

In WO 97/18325 (ref. 2), a method for determining the presence of specific nucleic acid sequences in a sample of eukaryotic origin using *in situ* hybridisation is disclosed. The method comprises the steps of producing a preparation of the sample to be investigated, contacting the preparation with a hybridisation solution comprising at least one peptide nucleic acid probe and a hybrid destabilising agent, removing any unbound and non-specifically bound probe, and determining the presence of bound probe. The method may i.a. be used for detection of X chromosome specific sequences or centromeric sequences.

20 In comparison, the present invention is based on a different concept, namely the use of at least two sets of hybridisation probes capable of hybridising to sequences of at least two different regions of chromosomes related to the same potential chromosome aberration or in

25 different potential chromosome aberrations, thereby enabling determination of chromosome aberrations.

In DE 196 10 255 (ref. 3), nucleic acid sequences are disclosed, which sequences are specific for a translocation breakpoint of a chromosome. The sequences 35 comprise a first set of sequences complementary to a DNA

sequence of a first chromosome, which first set is capable of hybridising to the first DNA sequence, and a second set of sequences complementary to a DNA sequence of a second chromosome, which second set is capable of 5 hybridising to the second DNA sequence. Only nucleic acid probes are mentioned. The use of peptide nucleic acid probes is not mentioned, nor anticipated.

EP 727 487 (ref. 4) and EP 825 198 (ref. 5) relate to 10 Multi-tumour Aberrant Growth (MAG) genes and genes having a sequence of any one of the strands of any one of the members of the PLAG subfamily of zinc finger protein genes or CTNNB1 genes, respectively. Various methods for detecting chromosome aberrations are also described. The 15 concept of the methods described in these two documents is different from the concept of the present invention. The methods disclosed in EP 727 487 (ref. 4) and EP 825 198 (ref. 5), respectively, are based on a comparison between the aberrant gene and the wild-type gene, whereas 20 the method of the present invention enables direct detection of the potential chromosome aberration or aberrations. Furthermore, the use of peptide nucleic acid probes is not disclosed, nor anticipated.

25 DETAILED DESCRIPTION OF THE INVENTION

As mentioned above, the present invention relates to a method for determining the presence of chromosome aberrations in a sample of eukaryotic origin using *in situ* hybridisation, in which method at least two sets of hybridisation probes of peptide nucleic acid probes 30 capable of hybridising to sequences related to the potential aberration or aberrations are used.

The method is suitable for detecting sequences related to a broad range of potential aberrations, and thereby determining whether aberrations are present.

5 The sets of hybridisation probes may be used for determining nucleic acid sequences related to different chromosome aberrations or the same aberration.

Thus, in a particular embodiment of the present method,  
10 the potential aberration is a potential breakpoint.

Other examples of such potential aberrations are deletions, amplifications, inversions, duplications and aneuploidy.

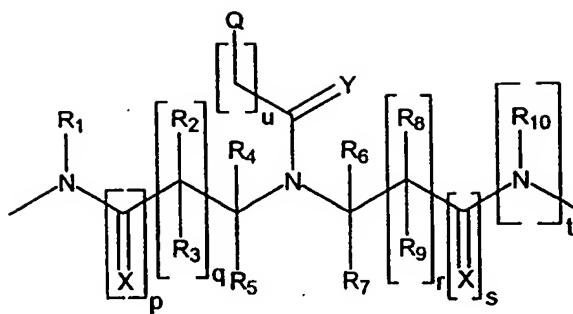
15 Furthermore, the present invention relates to a method for determining the presence of chromosome aberrations in a sample of eukaryotic origin using *in situ* hybridisation, in which method at least two sets of  
20 hybridisation probes of peptide nucleic acid probes flanking a potential breakpoint in a chromosome are used.

Suitable peptide nucleic acid probes to be used in the present invention are such containing from 8 to 40 polymerised moieties such as from 8 to 30 polymerised  
25 moieties.

30 Preferably, in the case of breakpoints, the sets of hybridisation probes should be chosen so as to hybridise to said specific nucleic acid sequences flanking a potential breakpoint in a chromosome within a genomic distance of no more than 100 kb, preferably no more than 50 kb from the potential breakpoint.

35 The probes are suitably such comprising polymerised moieties of formula (I)

5



(I)

wherein

10

Y designates O or S,

each X independently designates O or S,

each Q independently designates a ligand that is a naturally occurring nucleobase, a non-naturally occurring nucleobase, an intercalator, a nucleobase-binding group, a label or H,

u is an integer from 0, or 1 to 5,

p and s independently designate 0 or 1,

q and r independently designate 0 or 1,

20 t designates 0 or 1,

R<sub>1</sub> and R<sub>10</sub> independently designate H or C<sub>1-4</sub> alkyl,R<sub>2</sub>, R<sub>3</sub>, R<sub>4</sub>, R<sub>5</sub>, R<sub>6</sub>, R<sub>7</sub>, R<sub>8</sub> and R<sub>9</sub> independently designate H,

the side chain of a naturally occurring amino acid, or the side chain of a non-naturally occurring amino acid.

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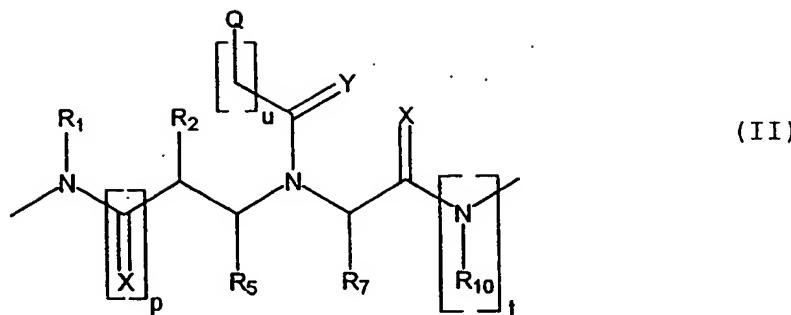
Suitable probes are such wherein Y, X, Q, u, p, q, r, s, R<sub>2</sub>, R<sub>5</sub>, R<sub>7</sub> and R<sub>8</sub> are as defined above, t is 0, R<sub>1</sub> designates H or CH<sub>3</sub>, and each of R<sub>3</sub>, R<sub>4</sub>, R<sub>6</sub> and R<sub>9</sub> designates H.

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Peptide nucleic acid probes comprising polymerised moieties of formula (II), which are moieties of the general formula (I) wherein r is 0 and q and s are 1,

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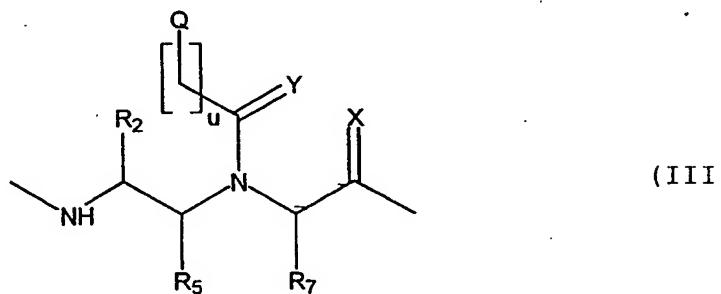
wherein Y, X, Q, p, t, u, R<sub>2</sub>, R<sub>5</sub> and R<sub>7</sub> are as defined in

claim 4, and

10 R<sub>1</sub> and R<sub>10</sub> independently designate H or CH<sub>3</sub>,  
are suitable for use in the present invention.

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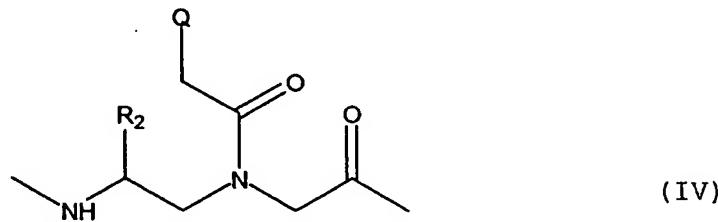
wherein Y, X, Q, u, R<sub>2</sub>, R<sub>5</sub> and R<sub>7</sub> are as defined above,  
may be used.

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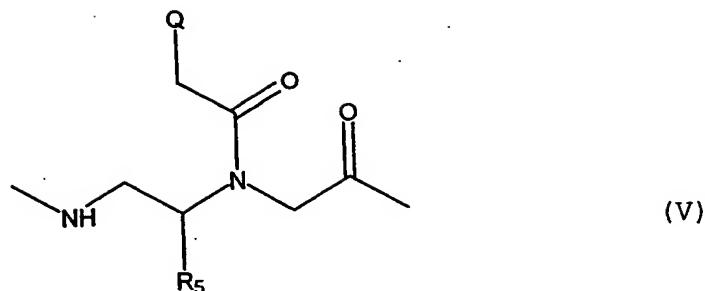
In a special embodiment, peptide nucleic acid probe comprising polymerised moieties of formulas (IV), (V) and/or (VI), which are moieties of the general formula (I) wherein p, r and t are 0, and u, s and q are 1,

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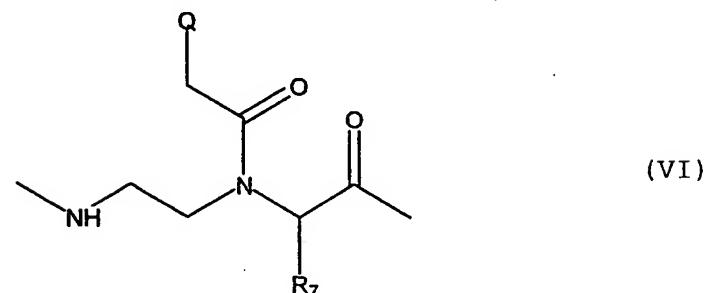
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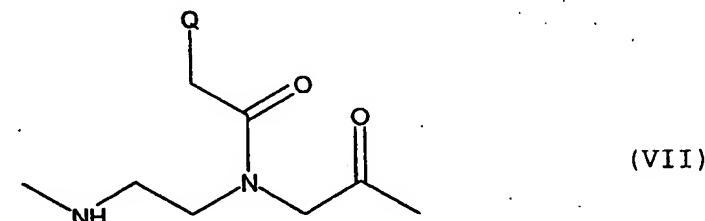
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wherein R<sub>2</sub>, R<sub>5</sub> and R<sub>7</sub> are as defined above, and each Q  
 15 independently designates a naturally occurring nucleobase, a non-naturally occurring nucleobase, an intercalator or a nucleobase-binding group, may be used. In particular, R<sub>2</sub>, R<sub>5</sub> and R<sub>7</sub> may designate H or the side chain of Ala, Asp, Cys, Glu, His, Homocys, Lys, Orn, Ser  
 20 or Thr, and Q designates thymine, adenine, cytosine, guanine, uracil or hypoxanthine.

Preferred peptide nucleic acid probe are those comprising polymerised moieties of formula (VII), which are moieties of formula (I) wherein p, r and t are 0, and u, s and q are 1

30



35 wherein Q designates thymine, adenine, cytosine, guanine, uracil or hypoxanthine.

The method of the invention can be used for detecting a broad range of chromosome aberrations, and further can be used for the diagnosis of disorders and diseases related 5 to chromosomal aberrations or abnormalities such as e.g. haemopoietic malignancies, cancers and inborn constituel diseases. Furthermore, the method may be used for detecting viral sequences and their localisation in the chromosome.

10

The term "chromosome aberration" is intended to mean translocation, amplification, inversion, duplication and aneuploidy.

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The term "nucleic acid sequences related to a potential aberration in a chromosome" is intended to comprise a region and regions of a chromosome/chromosomes of interest in relation to the aberration or aberrations in question.

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Examples of chromosome aberration-related disorders or diseases are acute lymphoblastic leukemias (ALL), chronic lymphotic leukemias, non-Hodgkin's lymphomas (NHL), malignant lymphomas, and multiple lymphomas. Well-

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established chromosome aberrations have been identified in many events. Often the particular chromosome aberration can be used to forecast the prognosis as some genetic aberrations are associated with favourable prognosis, whereas others are associated with poor 30 prognosis.

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Examples of other chromosome aberrations related to solid tumours and cancers are HER-2 (ERBB2) gene amplification frequently related to breast cancers. A breast cancer panel may also include e.g. CMYC (c-myc) amplification, EGFR (epidermal growth factor receptor) amplification or

duplication, amplification of genes localised to chromosome 20q13, TP53 (p53) deletion, and RB deletion. For other cancer types similar panels may be chosen. Also, some of the aberrations mentioned occur in other 5 cancer types.

Lymphoid malignancies are manifested in a broad range of diseases which differ in clinical symptoms, prognosis and treatment regimen. Such malignancies can be divided into 10 B-cell malignancies and T-cell malignancies. ALL can further can further be classified as pro-B-ALL, common-ALL, pre-B-ALL and additional T-ALL types.

In the case of ALL and NHL, several different types of 15 chromosome aberrations have been identified. For example, translocations resulting in fusion genes encoding for fusion proteins with new or modified functions have been identified in precursor-B-ALL (e.g. E2A-PBX and BCR-ABL fusion proteins from t(1;19) and t(9;22), respectively). 20 In acute leukemias, the 11q23 region with the MLL gene is important. Other examples are t(14;18) with involvement of the BCL2 gene, t(11;14) with involvement of the BCL1/Cyclin D1 gene, TAL1 deletions in T-ALL, t(12;21) in precursor-B-ALL, and t(2;5) in anaplastic large cell 25 lymphoma. Further examples are t(4;11) (q21;q23) involving the MLL-AF4 gene, t(11;19) (q23;p13) involving the MLL-ENL gene, t(6;11) (q27;q34) involving the MLL-AF6 gene, and t(9;11) (p22;q23) involving the MLL-AF9 gene in the case 30 of acute leukemias, t(2;18) (p12;q21) involving the IGK-BCL2 gene, and t(18;22) (q21;q11) involving the IGL-BCL2 gene in the case of malignant lymphomas, and t(11;22) (q24;q12) involving the EWS-FLI1 gene, t(21;22) (q22;q12) involving the EWS-ERG gene, and t(7;22) (p22;q12) involving the EWS-ETV1 gene in the case 35 of solid tumours.

In the method of the invention each peptide nucleic acid probe is suitably labelled directly or indirectly with at least one detectable label such as those selected from the group consisting of enzymes, chromophores, 5 fluorochromes, and haptens. Furthermore, various systems for enhancing or amplifying the signal may also be applied. Such systems are well-known in the art.

10 In a preferred embodiment of the invention labels are attached to the peptide nucleic acids in such a fashion that all peptide nucleic acids of one set of probes is labelled with one specific label while the peptide nucleic acids of the other set are labelled with another specific label.

15 For example, in the case of detection of breakpoints the use of different labels is an interesting option. By monitoring the fluorescence after hybridising the two sets of probes to the nucleic acid target to be 20 investigated the presence or absence of a breakpoint is readily apparent from the combined fluorescence of the peptide nucleic acid probe sets: if the two fluorescent signals are fused no break has occurred while the presence of two separate signals from each set of peptide 25 nucleic acid probes indicates, that a break has occurred between the target regions or targets investigated.

The method of the invention can be carried out using 30 various forms of detectable labels and detection methods. Also, the method of the invention may be utilised for simultaneous detection of several aberrations by employing several sets of peptide nucleic acid probes.

35 Peptide nucleic acids to be used according to the method of the invention can be selected on the basis of sequence analysis of nucleic acid regions to be investigated. The

peptide nucleic acids can be synthesised and labelled according to methods known in the art.

Hybridisation of the peptide nucleic acid sets to target  
5 material under investigation can be carried out for example as exemplified in the international patent application no. WO 97/18325 (ref. 2). Subsequent detection of the peptide nucleic acid hybridisation pattern is carried out according to methods well known in  
10 the art.

The hybrid destabilising agent is present in an amount effective to decrease the melting temperature of hybrids formed between the nucleic acid to be determined and the  
15 peptide nucleic acid probe used to detect said nucleic acid so as to increase the ratio between specific binding and non-specific binding. Within the present context, specific binding is intended to mean that the occurrence of false positive signals due to binding of the probes to  
20 other nucleic acid sequences having the same or similar sequence as the target sequence is reduced sufficiently to ensure determination under the used conditions.

Examples of hybrid destabilising agents are formamide,  
25 urea, ethylene glycol, guanidine and glycerol. Most of these agents may preferably be present in an amount of above 10% and less than 80%, such as 70%. Formamide and guanidine may more preferably be present in an amount of from 20% to 80%, more preferably of from 20% to 60%, most  
30 preferably of from 30% to 70% or from 30% to 50%. Urea is preferably used in a concentration from 2 to 5 M. Ethylene glycol may more preferably be present in an amount of from 30% to 65%, most preferably of from 50% to 65%.

The present invention provides a method for determining the presence of specific nucleic acid sequences in samples of eukaryotic origin, such as of human, animal or plant origin. Non-limiting examples are tissue sections, 5 cell smears, suspensions of cells or parts thereof and chromosome spreads. The presence of viral and/or bacterial DNA as well as non-eukaryotic DNA incorporated into eukaryotic DNA also may be detected using the present method.

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The peptide nucleic acid probe may suitably be labelled or unlabelled. In one aspect, the peptide nucleic acid probe carries one or more labels for detection of hybrids formed during hybridisation.

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In one embodiment of the present invention, a washing buffer at alkaline pH may preferably be used in step (c). The use of a washing buffer having an alkaline pH may in some cases provide an increased ratio between specific 20 binding and non-specific binding.

In another aspect, the present invention concerns a pair of set of hybridisation probes, one set hybridising to specific nucleic acid sequences related to a potential 25 aberration of a chromosome, and one set hybridising to specific nucleic acid sequences related to another or the same potential aberrations of a chromosome, each set comprising one or more peptide nucleic acid probes of formula (I) to (VII). In one embodiment, the pair of sets 30 is such, wherein the sets are capable of hybridising to nucleic acid sequences related to different chromosome aberrations. In another embodiment, the pair of sets is such which comprises sets of hybridisation probes capable of hybridising to different nucleic acid sequences 35 related to the same potential aberration. An example of

the latter is sets of probes capable of hybridising to nucleic acid sequences related to a potential breakpoint.

5 The invention further relates to a pair of sets of hybridisation probes, one set hybridising to specific nucleic acid sequences flanking one side of a potential breakpoint in a chromosome and another set hybridising to specific nucleic acid sequences flanking the other side of the potential breakpoint and each set comprising one  
10 or more peptide nucleic acid probes of formula (I) to (VII). The pair of sets of hybridisation probes is preferably such wherein each set hybridises to specific nucleic acid sequences flanking either side of a potential breakpoint in a chromosome within a genomic  
15 distance of no more than 100 kb, preferably no more than 50 kb from the potential breakpoint.

20 The pair of sets of hybridisation probes according to the invention is preferably such in which each set comprises from 1 to 500, from 1 to 250, from 1 to 100, from 1 to 35 peptide nucleic acid probes.

25 The invention also relates to an *in situ* diagnostic method for diagnosing chromosome aberrations in a sample of eukaryotic origin comprising the steps of

- a) producing a preparation of the sample, whereby the sample will be subject to a fixation,
- 30 b) contacting said preparation with a hybridisation solution comprising at least two sets of hybridisation probes, at least one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences related to a potential aberration in a chromosome, and at least another set comprising one or more

peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences related to another or the same potential aberration in a chromosome, and optionally a hybrid destabilising agent in an amount effective to decrease the melting temperature of hybrids formed between said nucleic acid sequences and said peptide nucleic acid probes so as to increase the ratio between specific and non-specific binding,

10

- c) removing any unbound and any non-specifically bound peptide nucleic acid probe, and
- d) determining the presence of peptide nucleic acid probe in the preparation.

In particular, one embodiment of the diagnostic method is an *in situ* diagnostic method for diagnosing chromosome aberrations in a sample of eukaryotic origin comprising the steps of

- a) producing a preparation of the sample, whereby the sample will be subject to a fixation,
- 25 b) contacting said preparation with a hybridisation solution comprising at least two sets of hybridisation probes, at least one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences flanking one side of a potential breakpoint in a chromosome, and at least one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences flanking the other side of the potential breakpoint and optionally a hybrid destabilising agent in an amount effective to decrease the melting temperature

of hybrids formed between said nucleic acid sequences and said peptide nucleic acid probes so as to increase the ratio between specific and non-specific binding,

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- c) removing any unbound and any non-specifically bound peptide nucleic acid probe, and
- 10 d) determining the presence of peptide nucleic acid probe in the preparation.

In a further embodiment of the diagnostic method, the method concerns an *in situ* diagnostic method for diagnosing chromosome aberrations in a sample of 15 eukaryotic origin comprising the steps of

- a) producing a preparation of the sample, whereby the sample will be subject to a fixation,
- 20 b) contacting said preparation with a hybridisation solution comprising at least two sets of hybridisation probes, one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences flanking one side of a potential breakpoint in a chromosome, and one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences flanking the other side of the potential breakpoint and optionally a hybrid destabilising 25 agent in an amount effective to decrease the melting temperature of hybrids formed between said nucleic acid sequences and said peptide nucleic acid probes so as to increase the ratio between specific and non-specific binding,
- 30

- c) removing any unbound and any non-specifically bound peptide nucleic acid probe, and
- d) determining the presence of peptide nucleic acid

5 probe in the preparation.

The invention further concerns the use of a pair of sets  
10 of hybridisation probes as defined above for use in the  
method of the invention, and for use in the *in vitro*  
diagnostic method of the invention.

#### Starting material

The present method can be used to detect nucleic acid  
15 sequences in samples of eucaryotic origin. It is  
contemplated that the present method provides a valuable  
tool for analysing such samples for the presence of  
nucleic acid sequences specific for e.g. pathogenic  
bacteria or virus hence providing information for  
20 establishing a diagnosis. In human or animal pathology,  
the detection of chromosomal aberrations may provide  
clinically important information for diagnosing genetic  
disorders or diseases. In plant biology, it is further  
contemplated that the present method may be a valuable  
25 tool for monitoring the efficiency of transferring for  
example herbicide resistance genes to a plant or, like in  
human tissues, to establish specific synthesis of  
proteins (by detection of mRNA) in a given cell  
structure.

30 The term "sample of eukaryotic origin" includes, but is  
not limited to samples of human, animal or plant origin.  
In the present context, the term "sample" is intended to  
include, but is not limited to, human, animal or plant  
35 tissue sections, cell or tissue cultures, suspension of  
human, animal or plant cells or isolated parts thereof,

human or animal biopsies, blood samples, saliva, urine, cerebrospinal fluid, milk, excretions, secretions, swabs, faecal samples and aspirates.

5 Producing a preparation of the sample

The sample to be examined is pre-treated before the hybridisation step whereby a preparation of the sample is produced. The person skilled in the art will readily 10 recognise that the appropriate pre-treatment will depend on the type of sample to be examined. During the pre-treatment, the sample will be subject to a fixation.

Thus, in one embodiment of the method, the sample is 15 deposited onto a solid support. Techniques for depositing a sample onto the solid support will depend upon the type of sample in question and may include, for example, sectioning of tissue as well as smearing or cytocentrifugation of cell suspensions. Many types of 20 solid supports may be utilised to practice the present method. The use of such supports and the procedures for depositing samples thereon are known to those skilled in the art. Glass microscope slides are especially convenient. Glass microscope slides can be treated to 25 better retain the sample.

Prior to hybridisation, the sample is suitably pre-treated with various chemicals and/or enzymes to facilitate the subsequent reactions. The actual pre- 30 treatment will depend on the type of sample to be analysed. The preferred treatment is one which fixes and preserves the morphological integrity of the cellular matrix and of the nucleic acids within the cell as well as enables the most efficient degree of probe 35 penetration.

In producing a preparation of a tissue sample, the morphological integrity of a tissue and the integrity of the nucleic acids can be preserved by bringing the sample to a fixed stage either by means of chemical fixation or 5 freezing. When freezing is used for preservation of for instance a biopsy, the biopsy is typically frozen in liquid nitrogen. After freezing, the sample may appropriately be stored at -80 °C. Prior to the analysis 10 of the nucleic acid, the frozen sample is cut into thin sections and transferred to e.g. pre-treated slides. This can e.g. be carried out at a temperature of -20 °C in a cryostat. The biopsy or tissue sections may suitably be stored at -80 °C until use. Prior to hybridisation, the tissue section may be treated with a fixative, e.g. a 15 precipitating fixative such as acetone and/or the tissue section is incubated for a short period in a solution of buffered formaldehyde. Alternatively, the biopsy or tissue section can be transferred to a fixative such as buffered formaldehyde for 12 to 24 hours. Following 20 fixation, the tissue may be embedded in paraffin forming a block from which thin sections can be cut. Well prepared paraffin-embedded samples can be stored at room temperature for a period of years.

25 Prior to hybridisation, the tissue section is deparaffinated and re-hydrated using standard procedures.

Further permeabilisation may be necessary in order to ensure sufficient accessibility of the target nucleic 30 acid sequences to the peptide nucleic acid probe. The type of treatment will depend on several factors, for instance on the fixative used, the extent of fixation, the type and size of sample used and the length of the peptide nucleic acid probe. The treatment may involve 35 exposure to protease such as proteinase K, pronase or

pepsin, diluted acids, detergents or alcohols or a heat treatment.

In some cases, a pre-hybridisation step using a pre-  
5 hybridisation mixture very similar to the hybridisation solution but without the peptide nucleic acid probe or another pre-hybridisation solution without probe, might be useful in order to decrease non-specific binding of the peptide nucleic acid probe. The components of the  
10 pre-hybridisation mixture should be selected so as to obtain an effective saturation of sites in the tissue that might otherwise bind the peptide nucleic acid probe non-specifically.

15 For analysing a suspended preparation such as a suspension of cells, the sample is treated so as to obtain a permeabilisation of the material and a preservation of the morphology. Fixation may be carried out with a fixative such as formaldehyde, acetone or  
20 ethanol.

The present method permits the detection of a specific nucleic acid sequence in a chromosome. Such detection may for example be carried out using spreads of chromosomes  
25 in metaphase, where the target sequence may be located in any region of the chromosomes. A sample preparation may in this case comprise adding an agent such as colcemide to arrest the chromosomes of dividing cells in the metaphase of the mitosis. Subsequently, the sample may be  
30 treated with a hypotonic buffer. Following centrifugation, the chromosomes are treated with a fixative and spread on a slide. Immediately before or simultaneously with the hybridisation step, the preparation may be treated to separate the double-  
35 stranded target. This separation can be achieved by heating the preparation in the presence of a denaturing

agent such as formamide. The detection of a specific nucleic acid sequence in a chromosome may also be carried out using chromosomes in interphase, e.g. in tissue sections or in a suspension of cells. In such cases, the 5 specimens may be treated as described above for tissue sections or suspension of cells.

For all sample preparations, the nucleic acids are fixed in a morphological structure allowing hybridisation to be 10 carried out *in situ*. Thus, the nucleic acids are not extracted from the cellular material and the hybridisation is not carried out in solution.

Peptide nucleic acids to be used in the present method

15 The term "naturally occurring nucleobases" includes the four main DNA bases (i.e. thymine (T), cytosine (C), adenine (A) and guanine (G)) as well as other naturally occurring nucleobases (e.g. uracil (U) and hypoxanthine).

20 The term "non-naturally occurring nucleobases" comprises, e.g., naturally occurring nucleobases wherein a label is coupled to the base optionally through a suitable linker, and modified naturally occurring nucleobases such as, 25 e.g., modified uracil and hypoxanthine. Other examples of non-naturally occurring nucleobases are 2,6-diamino purine, propynylcytosine (C propynyl), isocytosine (iso-C), isopseudouracil (iso<sup>pseudo</sup>C), 5-methyl-isocytosine (iso<sup>Me</sup>C), and thiouracil (see e.g. Tetrahedron Letters 30 Vol 36, No 21, 3601-3604 (1995) (ref. 6)).

Examples of useful intercalators are e.g. acridin, antraquinone, psoralen and pyrene.

35 Examples of useful nucleobase-binding groups are e.g. 3-nitro pyrrole and 5-nitro indole.

In the present context, "C<sub>1-4</sub> alkyl" is intended to mean branched or non-branched alkyl groups containing 1 to 4 carbon atoms. Non-limiting examples are CH<sub>3</sub>, CH<sub>2</sub>CH<sub>3</sub> and 5 CH(CH<sub>3</sub>)<sub>2</sub>.

Within the present context, the expression "naturally occurring amino acid" is intended to comprise D- and L-forms of amino acids commonly found in nature, e.g. D- 10 and L-forms of Ala (alanine), Arg (arginine), Asn (asparagine), Asp (aspartic acid), Cys (cysteine), Gln (glutamine), Glu (glutamic acid), His (histidine), Ile (isoleucine), Leu (leucine), Lys (lysine), Met (methionine), Phe (phenylalanine), Pro (proline), Ser (serine), 15 Thr (threonine), Trp (tryptophan), Tyr (tyrosine) and Val (valine).

In the present context, the expression "non-naturally occurring amino acid" is intended to comprise D- and L-forms of amino acids other than those commonly found in 20 nature as well as modified naturally occurring amino acids. Examples of useful non-naturally occurring amino acids are D- and L-forms of Cha (cyclohexylalanine), Cit (citrulline), Hci (homocitrulline), HomoCys (homocysteine), Hse (homoserine), Nle (norleucine), Nva (norvaline), Orn (ornithine), Sar (sarcosine) and Thi (thienylalanine).

The peptide nucleic acid probes to be used in the present 30 method should comprise a sufficient number of ligands capable of interacting with the nucleotide sequence to be determined to form hybrids sufficiently stable under the stringency used in the hybridisation and in the post-hybridisation wash. The strategy for selecting the 35 ligands of the peptide nucleic acid probe to be used in

accordance with the present method may be based on available target nucleotide sequence information.

In the above-indicated peptide nucleic acid probes, the 5 backbone of the moieties may preferably consist of six atoms. This has been shown to provide the presently strongest observed affinity for nucleic acids. It may in some cases be advantageous to change the strength of the binding between the peptide nucleic acid probe and the 10 nucleic acid sequence. Such change of the affinity may be accomplished by separating the ligands by fewer or by more atoms than six atoms. It is contemplated that preferred peptide nucleic acid probes to be used in the present method comprise less than 25% by weight 15 (calculated excluding X and Q groups as well as any linkers and/or labels) of moieties having more or less than six atoms in the backbone.

The strength of the binding between the peptide nucleic 20 acid probe and the nucleic acid sequence is influenced by the ligand Q. Hoogsteen and/or Watson-Crick base pairing assist in the formation of hybrids between a nucleic acid and a peptide nucleic acid probe wherein Q designates a nucleobase. It is contemplated that one or more of the 25 ligands may be a group which contribute little or none to the binding of the nucleic acid such as hydrogen. It is contemplated that peptide nucleic acid probes to be used in the present method comprise less than 25% by weight moieties wherein Q designate H. One or more of the 30 ligands Q may be groups that stabilise nucleobase stacking such as intercalators.

In the above-indicated peptide nucleic acid probes, one or more of the Q-groups may designate a label. Examples 35 of suitable labels are given below. Moieties wherein Q denotes a label may preferably be located in one or both

of the terminating moieties of the peptide nucleic acid. Moieties wherein Q denotes a label may also be located internally.

5 The peptide nucleic acid comprises polymerised moieties as defined above. From the formula, it is to be understood that the peptide nucleic acid probe may comprise polymerised moieties the structure of which may be mutually different or identical.

10

The backbone of the peptide nucleic acid probe may suitably form a polyamide comprising polymerised moieties, wherein all X groups are O, or form a polythioamide comprising polymerised moieties, wherein 15 all X groups are S. The polyamide and polythioamide forming moieties may be linked to form probes comprising both polyamide and polythioamide forming moieties or to form probes comprising polyamide forming moieties or polythioamide forming moieties only. Peptide nucleic acid 20 probes having a polyamide backbone (all X groups designate O) are of particular interest.

The preferred length of the peptide nucleic acid probe will depend on the target material and whether labelled 25 peptide nucleic acid probes are used. It is contemplated that especially interesting peptide nucleic acid probes comprise from 8 to 40 polymerised moieties as defined above. Peptide nucleic acid probes comprising from 8 to 30, from 12 to 25, or from 12 to 20 polymerised moieties 30 may be of particular interest, and peptide nucleic acid probes comprising from 14 to 23, and from 14 to 20 moieties are of most interest.

As mentioned above, the polymerised moieties of the 35 peptide nucleic acid probe may be mutually different or identical. In some embodiments, the polymerised moieties

of formulas (IV)-(VII) constitute at least 75% by weight (calculated as defined above), preferably at least 80% by weight and most preferably at least 90% by weight of the probe.

5

The ends on the moieties terminating the peptide nucleic acid may be substituted by suitable substituents. The substituents may be chosen so as to form the free or acylated form of the terminating moiety. The substituents 10 may further be chosen so as to form an ester, an amide or an amine depending on the chemical nature of the terminating moiety. A terminating end may further be substituted by one or more labels, which labels may be incorporated end to end, i.e. so as to form a non- 15 branched labelled end, or may be incorporated so as to form a branched labelled end ("zipper"). A terminating end may further be substituted by one or more linker units. Such linker units may be attached directly to a terminating end, may be attached to a label or between 20 labels on a terminating end, or be attached to a terminating end before a label is attached to a terminating end. It should be understood that two terminating ends may carry different or identical substituents, linker units and/or labels. It should 25 further be understood that the term "a label" is intended to comprise one or more labels.

The expression "peptide label" is intended to mean a label comprising from 1 to 20 naturally occurring or non- 30 naturally occurring amino acids, preferably from 1 to 10 naturally occurring or non-naturally occurring amino acids, more preferably from 1 to 8 naturally occurring or non-naturally occurring amino acids, most preferably from 1 to 4 naturally occurring or non-naturally occurring 35 amino acids, linked together end to end in a non-branched or branched ("zipper") fashion. In a preferred

embodiment, such a non-branched or branched end comprises one or more, preferably from 1 to 8 labels, more preferably from 1 to 4, further labels other than a peptide label. Such further labels may suitably terminate 5 a non-branched end or a branched end. One or more linker units may suitably be attached to the terminating end before a peptide label and/or a further label is attached. Such linker units may also be attached between a peptide label and a further label.

10

The probe as such may also comprise one or more labels such as from 1 to 8 labels, preferably from 1 to 4 labels, and/or one or more linker units, which may be attached internally, i.e. to the backbone of the probe. 15 The linker units and labels may mutually be attached as described above.

In the present context, the term "label" refers to a 20 substituent which is useful for detection of hybrids formed between a peptide nucleic acid probe and a nucleic acid. In accordance with the present invention, suitable labels comprise fluorophores, biotin, dinitro benzoic acid, digoxigenin, radioisotope labels, peptide or enzyme labels, dyes, chemiluminiscence labels, hapten, antigen 25 or antibody labels, and spin labels. Examples of particular interesting labels are biotin, fluorescent labels, such as fluorescein labels, e.g. 5-(and 6)-carboxyfluorescein, 5- or 6-carboxyfluorescein, 6-(fluorescein)-5-(and 6)-carboxamido hexanoic acid and 30 fluorescein isothiocyanate, peptide labels, dinitro benzoic acid, rhodamine, tetramethylrhodamine, cyanine dyes such as Cy2, Cy3 and Cy5, optionally substituted coumarin, R-phycoerythrin, allophycoerythrin, Texas Red and Princeton Red as well as conjugates of R-35 phycoerythrin and, e.g. Cy5 or Texas Red.

Examples of preferred labels are biotin, fluorescent labels, peptide labels and dinitro benzoic acid. Peptide labels may preferably be composed of from 1 to 10, more preferably of from 1 to 8, most preferably of from 1 to 5, naturally occurring or non-naturally occurring amino acids. It may be particularly advantageous to incorporate one or more other labels as well as a peptide label such as from 1 to 8 or from 1 to 4 other labels. Two of such other labels may e.g. be incorporated at each terminating 10 end.

Suitable peptide labels may preferably be composed of cysteine, glycine, lysine, ornithine, glutamic acid or aspartic acid.

15 A linker may be made up of linker units selected from units of formulas  $-\text{NH}(\text{CH}_2\text{CH}_2\text{O})_n\text{CH}_2\text{C}(\text{O})-$ ,  $-\text{NH}(\text{CHOH})_n\text{C}(\text{O})-$ ,  $-(\text{O})\text{C}(\text{CH}_2\text{OCH}_2)_n\text{C}(\text{O})-$  and  $-\text{NH}(\text{CH}_2)_n\text{C}(\text{O})-$ , wherein n is 0 or an integer from 1 to 8, preferably from 1 to 3. A linker 20 unit may have a free amino group or a free acid group, i.e.  $\text{NH}_2(\text{CH}_2\text{CH}_2\text{O})_n\text{CH}_2\text{C}(\text{O})-$ ,  $\text{NH}_2(\text{CHOH})_n\text{C}(\text{O})-$ ,  $\text{HO}(\text{O})\text{C}(\text{CH}_2\text{OCH}_2)_n\text{C}(\text{O})-$ ,  $\text{NH}_2(\text{CH}_2)_n\text{C}(\text{O})-$ ,  $-\text{NH}(\text{CH}_2\text{CH}_2\text{O})_n-$   $\text{CH}_2\text{C}(\text{O})\text{OH}$ ,  $-\text{NH}(\text{CHOH})_n\text{C}(\text{O})\text{OH}$ ,  $-(\text{O})\text{C}(\text{CH}_2\text{OCH}_2)_n\text{C}(\text{O})\text{OH}$  and  $-\text{NH}(\text{CH}_2)_n\text{C}(\text{O})\text{OH}$ . A linker may consist of up to 3 of such 25 linker units. Examples of very interesting linker units are  $-\text{NHCH}_2\text{C}(\text{O})-$ ,  $-\text{NHCH}_2\text{CH}_2\text{C}(\text{O})-$ ,  $-\text{NH}(\text{CH}_2\text{CH}_2\text{O})_2\text{CH}_2\text{C}(\text{O})-$ ,  $\text{HO}(\text{O})\text{CCH}_2\text{C}(\text{O}) (\text{NH}(\text{CH}_2\text{CH}_2\text{O})_2\text{CH}_2\text{C}(\text{O}))_2-$ .

30 In a further embodiment, the ligand Q as defined above may be labelled. Suitable labels are as defined above. Between such a label, a linker selected from  $\text{C}_{1-15}$  alkyl,  $\text{C}_{1-15}$  alkenyl and  $\text{C}_{1-15}$  alkynyl may be incorporated. In the present context, " $\text{C}_{1-15}$  alkyl,  $\text{C}_{1-15}$  alkenyl and  $\text{C}_{1-15}$  alkynyl" are intended to mean branched or non-branched 35 alkyl, alkenyl and alkynyl groups containing from 1 to 15 carbon atoms. It is preferred that such labelled ligands

Q are selected from thymine and uracil labelled in the 5 position.

In a preferred embodiment, probes used are peptide 5 nucleic acid probes comprising polymerised N-(2-aminoethyl)glycine moieties of formula (VII) wherein the glycine nitrogen is connected to naturally occurring nucleobases by a methylene carbonyl linker. The peptide nucleic acid probes to be used as detection probes may 10 suitably comprise from 8 to 40, from 8 to 30 of such polymerised moieties, preferably from 12 to 25 or 12 to 20 moieties, most preferably from 14 to 23 or from 14 to 20 moieties.

15 Nucleobase sequence of the peptide nucleic acid probes

The nucleobase sequence of the peptide nucleic acid probes can be obtained by analysing the nucleobase sequence of the target sequence.

20 The nucleobase sequence of interesting genes can e.g. be obtained from the GenBank Database, or the information can be obtained by sequencing interesting genes.

25 The nucleobase sequence of each set of hybridisation probes is selected having regard to the following: Minimal degree of hairpin formation should be possible, minimal degree of self dimer formation should occur, and minimal degree of pair dimer formation should occur.  
30 Prospected probe sequences are compared with sequences obtained from relevant data bases like the ones available via URL:<http://www.NCBI.NLM.NIH.gov>.

35 Preparation of peptide nucleic acid probes for hybridisation

The peptide nucleic acid probes used in the examples were synthesised according to the procedures described in "PNA Information Package" obtained from Millipore Corporation (Bedford, MA, USA).

5

If using the Fmoc strategy for elongation of the peptide nucleic acid probe with linkers or amino acids, it was possible to have side chain amino groups protected with acid sensitive protection groups such as the Boc, the Mmt 10 or the Mtt group.

One way of labelling the peptide nucleic acid probe is to use a fluorescent label, such as 5-(and 6)-carboxy-fluorescein, 5- or 6-carboxyfluorescein, 6-(fluorescein)- 15 5-(and 6)-carboxamido hexanoic acid or fluorescein isothiocyanate. The acid group is activated with HATU (O-(7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexa-fluorophosphate) or HBTU (2-(1H-benzotriazol-1-yl)- 1,1,3,3-tetramethyluronium hexafluorophosphate) and 20 reacted with the N-terminal amino group or other side-chain or linker amino group. The same technique can be applied to other labelling groups containing an acid function. Alternatively, the succinimidyl ester of the above-mentioned labels may be used directly.

25

After synthesis, the peptide nucleic acid probes are cleaved from the resin using standard procedures as described by Millipore Corporation or PerSeptive Biosystems. The peptide nucleic acid probes can be 30 purified and analysed using reversed-phase HPLC techniques at 50°C and were characterised by matrix-assisted laser desorption/ionisation time of flight mass spectrometry (MALDI-TOFMS), plasma desorption mass spectrometry (PDMS) or electron spray mass spectrometry 35 (ESMS).

Generally, peptide nucleic acid probes such as peptide nucleic acid probes comprising polymerised moieties of formula (IV)-(VII) may also be prepared as described in, e.g., Tetrahedron Letters Vol 35, No 29, 5173-5176 (1994) (ref. 7). Chemical properties of some peptide nucleic acid probes are described in, e.g., Nature, 365, 566-568 (1993) (ref. 8).

#### Hybridisation

10

The hybridisation can be performed using fixed, immobilised or suspended preparations which are prepared as described above. If double-stranded target such as chromosomal or other DNA sequences are to be detected, a 15 treatment to separate the two strands may be necessary. This separation of the strands can be achieved by heating the sample in the presence of the hybridisation mixture to a temperature sufficiently high and for a time period sufficiently long to dissociate the strands. Typically, 20 heating at a temperature of 70°C to 100°C for a period of 1 to 15 minutes is suitable.

The hybridisation buffer may optionally comprise a hybrid destabilising agent in an amount effective to decrease 25 the melting temperature of hybrids formed between the nucleic acid to be determined and the peptide nucleic acid probe so as to increase the ratio between specific binding and non-specific binding. This agent will allow the hybridisation to take place at a lower temperature 30 than without the agent. In traditional nucleic acid hybridisation, such agent is called a denaturing agent.

The effective amount of the hybrid destabilising agent will depend on the type of destabilising agent used and 35 furthermore on the peptide nucleic acid probe or combination of peptide nucleic acid probes used. It has

been found that using peptide nucleic acid probes of formula (VII), particular good results have been obtained by including a hybrid destabilising agent. Examples of hybrid destabilising agents are formamide, urea, 5 guanidine, ethylene glycol and glycerol, and most of these agents may preferably be used in a concentration above 10% and less than 80%, such as 70%. Formamide and guanidine may more preferably be present in an amount of from 20% to 80%, more preferably of from 20% to 60%, most 10 preferably of from 30% to 70% or from 30% to 50%. Urea is preferably used in a concentration of from 2 to 5 M. Ethylene glycol may more preferably be present in an amount of from 30% to 65%, most preferably of from 50% to 65%.

15 It is often advantageous to include macromolecules or polymers such as dextran sulphate, polyvinylpyrrolidone and/or ficoll. In the presence of such macromolecules or polymers, the effective concentration of the peptide 20 nucleic acid probe at the target is assumed to be increased. Dextran sulphate may be added in a concentration of up to 15%. Concentrations of dextran sulphate of from 2.5% to 12.5% are often advantageous.

25 In some cases, it may be advantageous to add a detergent such as sodium dodecyl sulphate, Tween 20® or Triton X-100®.

30 During hybridisation, other important parameters are hybridisation temperature, concentration of the peptide nucleic acid probe and hybridisation time. The person skilled in the art will readily recognise that optimal conditions for various starting materials will have to be determined for each of the above-mentioned parameters.

The hybridisation between peptide nucleic acid probes and a target nucleic acid sequence appears to be less sensitive than DNA probes to variations in pH and in the concentration of NaCl.

5

#### Post-hybridisation washing

Following hybridisation, the preparation is washed to remove any unbound and any non-specifically bound peptide 10 nucleic acid probes. The conditions described below are merely by way of example and may depend on the type of preparation to be analysed. During the post-hybridisation step, appropriate stringency conditions should be used in order to remove any non-specifically bound peptide 15 nucleic acid probe. Stringency refers to the degree to which reaction conditions favour the dissociation of the formed hybrids and may be enhanced, for instance by increasing the washing temperature and washing time. For conventional hybridisation with nucleic acid probes, the 20 salt concentration is often used as an additional factor for regulating the stringency.

Examples of useful buffer systems are Tris-Buffered-Saline (TBS), standard citrate buffer (SSC) or phosphate 25 buffers. A convenient TBS buffer is 0.05 M Tris/HCl, 0.15 M NaCl, pH 7.6. The SSC buffer comprises 0.15 M sodium chloride and 0.015 M trisodium citrate, pH 7.0. Typically, washing buffers include a detergent such as sodium dodecylsulphate, Tween 20<sup>®</sup>, or Triton X-100<sup>®</sup>.

30

Typically, washing times from 5 to 30 minutes may be suitable. Washing periods of two times 10 minutes or 3 times 5 minutes in a suitable buffer may also give good results.

35

In some cases, particularly when using peptide nucleic acid probes carrying at least one fluorescein label, it has been shown to be advantageous to increase the pH of the washing buffer. In such cases, it is preferred that 5 the washing solution in step (c) has a pH value of from 8 to 10.5, preferably from 9 to 10.

Detection of hybrids formed

10 In cases where the preparation is deposited onto slides, the hybridisation results may be visualised using well known immunohistochemical staining methods to detect the labelling on the peptide nucleic acid probe. When 15 fluorescent labelled peptide nucleic acid probes are used, the hybrids may be detected using an antibody against the fluorescent label which antibody may be conjugated with an enzyme. The fluorescent label may alternatively be detected directly using a fluorescence microscope, or the results may be automatically analysed 20 on a fluorescent-based image analysis system.

When biotin labelled peptide nucleic acid probes are used, the hybrids may be detected using enzyme labelled streptavidin or antibodies against the biotin label which 25 antibody may be conjugated with an enzyme. If necessary, an enhancement of the signal can be generated using commercially available amplification systems such as the catalysed signal amplification system for biotinylated probes (DAKO K 1500). Digoxigenin labelled probes may be 30 detected using antibodies against digoxigenin, and dinitro benzoic acid with antibodies against dinitro benzoic acid.

In the case of a suspended sample such as a cell 35 suspension, quantitative results may be obtained using peptide nucleic acid probes comprising a fluorescent

label and a flow cytometer to record the intensity of fluorescence per cell. Signal amplification may be relevant for flow as well.

5 Peptide nucleic acid probes used in some aspects of the present method may form nucleic acid/peptide nucleic acid probe hybrids which can be recognised by an antibody described in WO 95/17430 (ref. 9). Hybrids formed between the peptide nucleic acid probe, nucleic acid and the 10 antibody can be detected in a direct immunohistochemical staining method using, for instance an enzyme conjugated form of the antibody, followed by detection of the enzyme activity or by the application of well known indirect immunohistochemical staining techniques.

15

#### ILLUSTRATIVE EMBODIMENTS

##### ILLUSTRATIVE EMBODIMENT 1

20 The method of the invention can suitably be used for detecting chromosome aberrations in ALL and NHL. The sets of hybridisation probes are chosen so as to flank each site of the potential breakpoint. Interesting regions are indicated below in Tables 1 and 2. Apart from the two 25 sets flanking the potential breakpoint, at least one set of probes targeting fusion partner genes may be included

TABLE 1. Chromosome aberrations in ALL.

Type of ALL	Chromosome aberration	
	Type	Involved genes
Precursor-B-ALL	t(1;19) (q23;p13)	E2A-PBX1
	t(4;11) (q21;q23)	MLL-AF4
	t(9;22) (q34;q11)	BCR-ABL
	t(12;21) (p13;q22)	TEL-AML1
T-ALL	TAL1 deletion	SIL-TAL1
	t(1;14) (q34;q11)	TAL1-TCRD
	t(10;14) (q24;q11)	HOX11-TCRD
	t(11;14) (p13;q11)	RHOM2-TCRD

TABLE 2. Chromosome aberrations in NHL.

Type of NHL	Chromosome aberration	
	Type	Involved genes
Follicular NHL	t(14;18) 3q37	IGH-BCL2 BCL6
Diffuse large cell centroblastic NHL	t(14;18) 3q27	IGH-BCL2 BCL6
Large cell anaplastic NHL	t(2;5)	NPM-ALK
Mantle cell NHL	t(11;15)	BCL1-IGH
Immunoblastair NHL	t(14;18)	IGH-BCL2
Burkitt's NHL	t(8;14) t(2;8) t(8;22)	MYC-IGH IGK-MYC IGL-MYC
Lymphocytic NHL		
Immunocytoma		
MALT-NHL		
Peripheral T-NHL		
Lymphoblastic T-NHL	TAL1 deletion t(1;14) t(10;14) t(11;14)	SIL-TAL1 TAL1-TCRD HOX11-TCRD RHOM1-TCRD

## ILLUSTRATIVE EMBODIMENT 2

5

Determination of translocation involving the major breakpoint (mbr) in the MLL gene (ALL-1) and HRX

10 The nucleobase sequences of the peptide nucleic acid probes are selected from EMBL + GenBank, Accession numbers HSALL1X28 (exon 28) and HSALL1X4 (exon 4). The nucleobases are chosen so as to form two sets of probes, a set flanking each site of the mbr region, e.g. from

exon 4 and exon 28. Each set of hybridisation probes consists suitably of from 1 to 500, from 1 to 250, from 1 to 100, or from 1 to 35 peptide nucleic acid probes.

5 The nucleobase sequence of the probes indicated below are selected from the lower strand (i.e. they hybridise to the upper strand). The position of the nucleobases relative to the whole target sequence are indicated as "position". The first number corresponds to the position 10 of the 5' end of the probe in the sequence, and the last number corresponds to the position of the 3' end of the probe.

Two suitable sets of probes are e.g.

15

	<u>ALL-1 EXON 4 (one set)</u>	<u>Position</u>	
	GGGAAACACAGATGGAT	120-103	Seq ID No 1
	GATTTGGTCTCTGATTTATTTAG	145-123	Seq ID No 2
	GTTATTTGATTTTACTCCAG	235-214	Seq ID No 3
20	GGTAACCTGAAATGTCCTT	243-262	Seq ID No 4
	TTTTTCAGGCTATCTTCTT	292-274	Seq ID No 5
	CTTAAACTTAGACTTGAGAGGA	386-365	Seq ID No 6
	CCCCTTCCTTCCTATT	416-400	Seq ID No 7
	CCTTCCTCTCCGTCGTA	443-427	Seq ID No 8
25	GGGTCTTATCCTTCTGT	471-453	Seq ID No 9
	ACTTTCTGGGGCTTTTC	517-501	Seq ID No 10
	TGTTCCCTCCTTGTCTTTC	539-521	Seq ID No 11
	AACTGTCTTATCTTCTTTGTA	569-548	Seq ID No 12
	CTTCGAGGGCTTGTCT	589-573	Seq ID No 13
30	GCATCTGTCCTTTGAA	634-617	Seq ID No 14
	CTTCTTTTCAATTTCCTTT	699-679	Seq ID No 15
	GGGGCACTGAATCTACTAT	889-871	Seq ID No 16
	AGCTGCACTGATTTTC	923-906	Seq ID No 17
	CGAATATTTTGACCTGTG	748-730	Seq ID No 18
35	GGTCATAATCCTCATCCTC	834-816	Seq ID No 19
	AGGAGATAGCACTGACAACA	783-764	Seq ID No 20

TTTGAGAGGAGTGCTGAGA	942-924	Seq ID No 21
TATCAACACTGGGGCTACTA	965-984	Seq ID No 22

<u>ALL-1 EXON 28 (another set)</u>		<u>Position</u>	
5	TACTGTGACGCCCTGGAG	123-107	Seq ID No 23
	AGTGGTTGGGGTAGGAC	50-34	Seq ID No 24
	GCTGGGGTGATAAGGAA	144-128	Seq ID No 25
	CGGTGGTGGAGACTGA	225-210	Seq ID No 26
	TTGAAGAGGTGGAAGTGT	327-309	Seq ID No 27
10	CACTGGACTGCTTCATTTC	411-393	Seq ID No 28
	ACTCAGCAGTTGGTCTTC	470-452	Seq ID No 29
	AAGGAGCCGATTTACTA	569-586	Seq ID No 30
	GAGGTGTGGGAAGGAGA	644-628	Seq ID No 31
	GTTCGCTGATTGGTAGAA	704-686	Seq ID No 32
15	ATGTTCGTTGATAATGGAT	785-767	Seq ID No 33
	CTTCCTCACCAAGTTGTCAC	906-888	Seq ID No 34
	CAAAACCTCATCCATAAAC	941-923	Seq ID No 35
	GAACCTGGCATAGAAAG	1025-1008	Seq ID No 36
	TTCCGTGGTGCCTGTA	1084-1069	Seq ID No 37
20	CCATTTTAGAGGTGTCAG	1122-1104	Seq ID No 38
	CATTTTGGATTGACTCTC	1146-1128	Seq ID No 39
	GCAGGACTACTTCTTTCA	1168-1150	Seq ID No 40
	CTGTGGGAGATGTTGACT	1203-1186	Seq ID No 41
	TATTATTGGGGCTTGGTT	1266-1249	Seq ID No 42

25

The probes can of course be selected from the upper strand (i.e. so as to be capable to hybridise to the lower strand).

30 The probes are preferably labelled. E.g. the set of probes for exon 4 may be labelled with one or more fluorescein labels, and the set of probes for exon 28 with one or more Cy3.

35 Step (a): Preparation of sample and pre-treatment

Metaphase chromosome spreads can be prepared from peripheral blood, bone marrow, amnion, chorion villus samples or tumour cells. If prepared from peripheral blood, a sample of whole blood collected in a glass tube 5 containing heparin or another anticoagulant is mixed with suitable growth medium. As an Example, 0,5 ml whole blood may be added to 10 ml RPMI 1640 medium supplemented with 20% (v/v) foetal calf serum, 2 mM glutamine, 100 U penicillin/streptomycin, 1% phytohemagglutinin and 5 U/ml 10 heparin. The sample is incubated in a CO<sub>2</sub> incubator, e.g. at 37°C for 72 h. The chromosomes are arrested in the metaphase of the mitosis, e.g. by adding colcemid (0.1 µg/ml) to the culture approximately 1½ hours before harvesting the cells.

15

The cells are collected by centrifugation and re-suspended in a hypotonic buffer, e.g. 60 mM KCl at room temperature for about 30 minutes. The sample is treated with a suitable fixative such as methanol:acetic acid 20 3:1. The treatment with fixative may be repeated several times. After treatment the suspensions are left at freezing temperatures such as at -20°C from 20 minutes to 3 days. Chromosome spreads are prepared by spotting a suitable amount of the suspension onto a clean slide and 25 air-drying the slide.

Prior to hybridisation it may be advantageous to submit the specimens on the slides to a pre-treatment including RNase treatment and/or additional fixation and/or 30 protease digestion. The following procedure may e.g. be applied:

- 1) Immerse slides in TBS 2 min.
- 2) Immerse slides in 3,7% formaldehyde in TBS for 2 min.
- 35 3) Wash slides 3x5 min. in TBS.

- 4) Immerse slides in proper dilution of Proteinase K for 10 min at room temperature.
- 5) Incubate slides in RNase 10-100 $\mu$ g/ml in 2xSSC for 20-60 min. at 37°C.
- 5) Wash 3x5 min. in TBS.
- 7) Dehydrate specimens in cold ethanol series (70%, 85% and 96%) and air dry them.

Step (b): Hybridisation

10

Prior to hybridisation, the chromosomal DNA is denatured. Such denaturing may be carried out by placing the slide in a solution of 2xSSC and 70% formamide at about 80°C for a short period of time such as 3-5 minutes.

15

Denaturation may be performed by adding the probes in proper dilution in the hybridisation buffer onto the slide containing the sample and covering it with a coverslip. The slides are then placed in a preheated 20 incubator adjusted to 80°C for 3 min. Hybridisation may be performed by placing the slide in the dark at room temperature or higher temperatures (50-60°C may be suitable) for 5-60 min.. A suitable hybridisation buffer is 10 mM Tris pH 7.2 with 70% formamide containing 25 conventional blocking reagent.

Step (c): Removal of any unbound and any non-specifically bound peptide nucleic acid probe

30 The slide may be rinsed in TBS, pH 7.4 or 0,1xSSC to remove the coverslips followed by wash for about 5-45 min. in an appropriate washing buffer e.g. 0.1 x SSC (15 mM NaCl, 1.5 mM sodium citrate pH 7.0) with 0.1% Triton X-100® or PBS with 0.1% Tween-20®.

35

Step (d): Detection of hybrids formed

If the bound probes are fluorescence labelled visible signals may be found by inspection of the slides in the fluorescence microscope at this step. Additional 5 amplification of the signal may be provided using some of the many different immunohistochemical/immunocytochemical amplification systems well known in the art. Non-fluorescent labelled probes may also be detected using some of the many different immunohistochemical/- 10 immunocytochemical amplification systems well known in the art.

Following appropriate amplification/detection, slides are mounted for inspection in the fluorescence microscope. A 15 mounting media may include counter stain such as DAPI. A successful hybridisation should preferably result in easily visible signals/spots on the chromosomes and within the interfase nuclei. There should preferably be no interfering non-specific hybridisation on the 20 chromosomes. The microscope should be correctly equipped for the labels used.

#### EXAMPLES

##### 25 EXAMPLE 1

###### Sequence of peptide nucleic acid probes

One set of probes targeting HER-2 was selected (Accession 30 number emb|X03363|HSERB2R Human c-erb-B-2 mRNA). The probes were selected and numbered as described in Illustrative embodiment 2. The nucleobase sequences of the peptide nucleic acid probes were as follows:

35		<u>Position</u>
	CAAGAGGGCGAGGGAGGAG	222-205
		Seq. ID No 43

	GGTAGAGGTGGCGGAGCAT	325-307	Seq ID No 44
	TGGGCAGGTAGGTGAGTTC	373-355	Seq ID No 45
	GTGGGCAGGTAGGTGAGTT	374-356	Seq ID No 46
	GGCCAGGGCATAGTTGTC	519-502	Seq ID No 47
5	GTATTGTTCAGCGGGTCTC	551-533	Seq ID No 48
	GCAGAGCTGGGGGTTCC	660-644	Seq ID No 49
	GAGCCAGCTGGTTGTTCT	715-698	Seq ID No 50
	GCGGTTGGTGTCTATCAGT	738-720	Seq ID No 51
	AGGCCAGGCAGTCAGAGTG	937-919	Seq ID No 52
10	CCACGTCCGTAGAAAGGTA	1099-1081	Seq ID No 53
	GCAGGGGGCAGACGAG	1126-1111	Seq ID No 54
	GGGCTTGCTGCACTTCTCA	1185-1167	Seq ID No 55
	GCAGCCAGCAAACCTCC	1275-1260	Seq ID No 56
	GAGCGGGGCAGTGTGGG	1350-1334	Seq ID No 57
15	GAGTAGGCGGCCATTGTGC	1499-1482	Seq ID No 58
	CTTGCAGGGTCAGCGAGTA	1513-1495	Seq ID No 59
	GGTGTATGGTGGATGAGG	1590-1572	Seq ID No 60
	AGCTTGGTGCAGGGTTCC	1650-1634	Seq ID No 61
	CCGGTTGGCAGTGTGGAG	1671-1654	Seq ID No 62
20	CCTGGCATTACACATACTCC	1848-1830	Seq ID No 63
	AGGTCACTGAGCCATTCTG	1900-1882	Seq ID No 64
	GGAGAGGTCAGGTTTCACA	2001-1983	Seq ID No 65
	GAGTGGGTGCAGTTGATGG	2072-2054	Seq ID No 66
	CACCGCAGAGATGATGGAC	2148-2130	Seq ID No 67
25	TGAGGATCCCAAAGACCAC	2197-2179	Seq ID No 68
	ATGCCCTTGTAGACTGTGC	2387-2369	Seq ID No 69
	AGCTGCACCGTGGATGTCA	2561-2543	Seq ID No 70
	GAGCCAGCCCGAAGTCT	2776-2760	Seq ID No 71
	CTTGGCCGACATTCAAGAGT	3077-3059	Seq ID No 72
30	TGGCCATGCGGGAGAAT	3115-3099	Seq ID No 73
	CAGTGAGCGGGTAGAAGGTG	3198-3180	Seq ID No 74
	CGGTGCCTGTGGTGGAC	3317-3301	Seq ID No 75
	CATCTGGCTGGTCACATA	3607-3589	Seq ID No 76
	TTGAAGGTGCTGGGTGGAG	3887-3869	Seq ID No 77
35	TCAAGCAGGAAGGAAGGTT	4099-4081	Seq ID No 78

All probes are labelled with one or two fluorescein labels. As linkers, one or two  $\text{NH}_2\text{CH}_2\text{CH}_2(\text{COCH}_2\text{OCH}_2\text{CON}(\text{CH}_2\text{CH}_2\text{OCH}_3)_2\text{CH}_2\text{COOH}$ -linkers and/or one or two  $\text{NH}_2-(\text{CH}_2\text{CH}_2\text{O})_2\text{CH}_2\text{C(O)}$ -linkers were used.

5

Another set of probes targeting the centromer of chromosome 17 was selected. These probes are included because the HER-2 gene in normal cells are located to chromosome 17, and aneuploidy is frequently observed in 10 cancer cells. The nucleobase sequences of the peptide nucleic acid probes were as follows:

	AACGAATTATGGTCACAT	Seq ID No 78
	GGTGACGACTGAGTTAA	Seq ID No 79
15	TTTGGACCACTCTGTGGC	Seq ID No 80
	AACGGGATAACTGCACCT	Seq ID No 81
	TTTGTGGTTGTGGTGGA	Seq ID No 82
	AGGGAATAGCTTCATAGA	Seq ID No 83
	ATCACGAAGAAGGTTCTG	Seq ID No 84
20	CCGAAGATGTCTTGAA	Seq ID No 85
	AAAGAGGTCTACATGTCC	Seq ID No 86

All probes were labelled with one or two fluorescein labels. As linkers, one or two  $\text{NH}_2\text{CH}_2\text{CH}_2(\text{COCH}_2\text{OCH}_2\text{CON}(\text{CH}_2\text{CH}_2\text{OCH}_3)_2\text{CH}_2\text{COOH}$ -linkers and/or one or two  $\text{NH}_2-(\text{CH}_2\text{CH}_2\text{O})_2\text{CH}_2\text{C(O)}$ -linkers were used.

The probes were synthesised according to standard PNA synthesis procedures.

30

Sample preparation, hybridisation, washing and detection are carried out as described above under Illustrative embodiment 2.

## REFERENCES

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2. WO 97/18325
- 5 3. DE 196 10 255
4. EP 727 487
5. EP 825 198
6. Tetrahedron Letters Vol. 36, No. 21, 3601-3604 (1995)
7. Tetrahedron Letters Vol. 35, No. 29, 5173-5176 (1994)
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9. WO 95/17430

## CLAIMS

1. A method of determining the presence of chromosome aberrations in a sample of eukaryotic origin using *in situ* hybridisation, comprising the steps of
  - 5 a) producing a preparation of the sample, whereby the sample will be subject to a fixation,
  - 10 b) contacting said preparation with a hybridisation solution comprising at least two sets of hybridisation probes, at least one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences
  - 15 related to a potential aberration in a chromosome, and at least one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences related to another or the same potential aberration in a chromosome, and
  - 20 optionally a hybrid destabilising agent in an amount effective to decrease the melting temperature of hybrids formed between said nucleic acid sequences and said peptide nucleic acid probes so as to increase the ratio between specific and non-specific
  - 25 binding,
- c) removing any unbound and any non-specifically bound peptide nucleic acid probe, and
- 30 d) determining the presence of peptide nucleic acid probe in the preparation.

2. A method according to claim 1, wherein each set of hybridisation probes hybridise to specific nucleic acid sequences related to the same potential chromosome aberration.

3. A method according to claim 1 or 2, wherein the potential aberration is a potential breakpoint.
- 5 4. A method of determining the presence of chromosome aberrations in a sample of eukaryotic origin using *in situ* hybridisation, comprising the steps of
  - 10 a) producing a preparation of the sample, whereby the sample will be subject to a fixation,
  - 15 b) contacting said preparation with a hybridisation solution comprising at least two sets of hybridisation probes, at least one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences flanking one side of a potential breakpoint in a chromosome, and at least one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences flanking the other side of the potential breakpoint and optionally a hybrid destabilising agent in an amount effective to decrease the melting temperature of hybrids formed between said nucleic acid sequences and said peptide nucleic acid probes so as to increase the ratio between specific and non-specific binding,
  - 25 c) removing any unbound and any non-specifically bound peptide nucleic acid probe, and
  - 30 d) determining the presence of peptide nucleic acid probe in the preparation.

5. A method of determining the presence of chromosome aberrations in a sample of eukaryotic origin using *in situ* hybridisation, comprising the steps of

5    a) producing a preparation of the sample, whereby the sample will be subject to a fixation,

10    b) contacting said preparation with a hybridisation solution comprising at least two sets of hybridisation probes, one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences flanking one side of a potential breakpoint in a chromosome, and one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences flanking the other side of the potential breakpoint and optionally a hybrid destabilising agent in an amount effective to decrease the melting temperature of hybrids formed between said nucleic acid sequences and said peptide nucleic acid probes so as to increase the ratio between specific and non-specific binding,

15    c) removing any unbound and any non-specifically bound peptide nucleic acid probe, and

20    d) determining the presence of peptide nucleic acid probe in the preparation.

30    6. A method according to claim 4 or 5, wherein each set of hybridisation probes hybridise to said specific nucleic acid sequences flanking a potential breakpoint in a chromosome within a genomic distance of no more than 100 kb, preferably no more than 50 kb from the potential

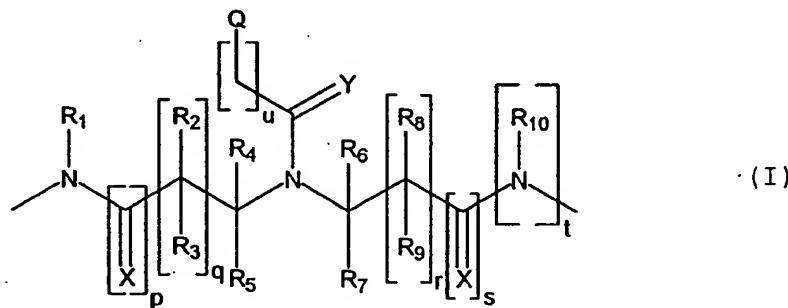
35    breakpoint.

7. A method according to any one of claims 1 to 6 wherein each peptide nucleic acid probe contains from 8 to 40 polymerised moieties.

5 8. A method according to any one of claims 1 to 7 wherein the peptide nucleic acid probe comprises polymerised moieties of formula (I)

10

15



wherein

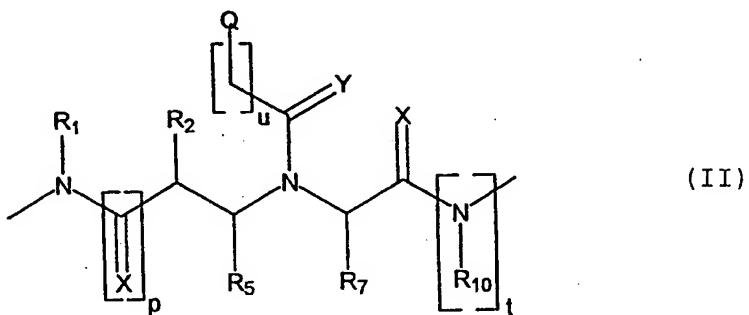
20 Y designates O or S,  
 each X independently designates O or S,  
 each Q independently designates a ligand that is a naturally occurring nucleobase, a non-naturally occurring nucleobase, an intercalator, a nucleobase-binding group,  
 25 a label or H,  
 u is an integer from 0, or 1 to 5,  
 p and s independently designate 0 or 1,  
 q and r independently designate 0 or 1,  
 t designates 0 or 1,  
 30 R<sub>1</sub> and R<sub>10</sub> independently designate H or C<sub>1-4</sub> alkyl,  
 R<sub>2</sub>, R<sub>3</sub>, R<sub>4</sub>, R<sub>5</sub>, R<sub>6</sub>, R<sub>7</sub>, R<sub>8</sub> and R<sub>9</sub> independently designate H, the side chain of a naturally occurring amino acid, or the side chain of a non-naturally occurring amino acid.  
 35 9. A method according to claim 8, wherein Y, X, Q, u, p, q, r, s, R<sub>2</sub>, R<sub>5</sub>, R<sub>7</sub> and R<sub>8</sub> are as defined in claim 8, t is

0,  $R_1$  designates H or  $CH_3$ , and each of  $R_3$ ,  $R_4$ ,  $R_6$  and  $R_9$  designates H.

10. A method according to claim 8, wherein the peptide  
5 nucleic acid probe comprises polymerised moieties of  
formula (II), which are moieties of the general formula  
(I) wherein r is 0 and q and s are 1,

10

15



wherein Y, X, Q, p, t, u,  $R_2$ ,  $R_5$  and  $R_7$  are as defined in  
claim 7, and

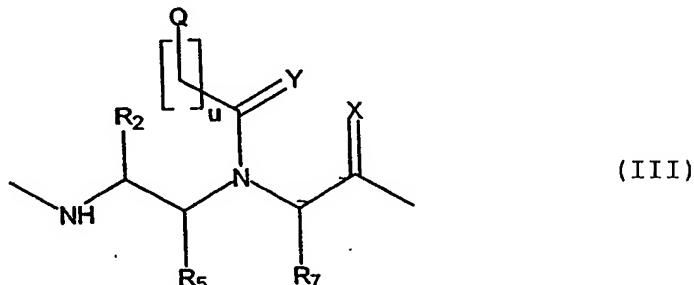
$R_1$  and  $R_{10}$  independently designate H or  $CH_3$ .

20

11. A method according to claim 8, wherein the peptide  
nucleic acid probe comprises polymerised moieties of  
formula (III), which are moieties of the general formula  
(I) wherein p, r and t are 0, and q and s are 1

25

30

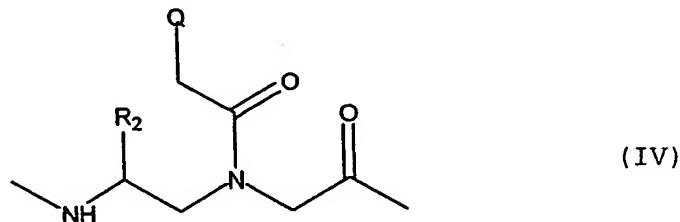


wherein Y, X, Q, u,  $R_2$ ,  $R_5$  and  $R_7$  are as defined in claim  
8.

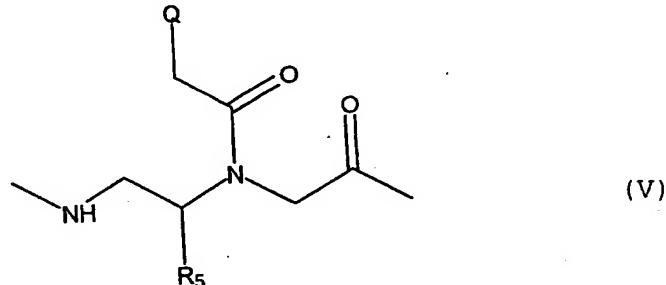
35

12. A method according to claim 8, wherein the peptide nucleic acid probe comprises polymerised moieties selected among those of formulas (IV)-(VI), which are moieties of the general formula (I) wherein p, r and t  
5 are 0, and u, s and q are 1,

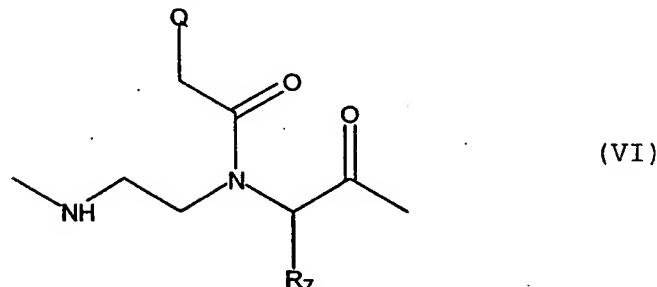
10



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wherein  $\text{R}_2$ ,  $\text{R}_5$  and  $\text{R}_7$  are as defined in claim 7, and each  $\text{Q}$  independently designates a naturally occurring nucleobase, a non-naturally occurring nucleobase, an intercalator or a nucleobase-binding group.

30

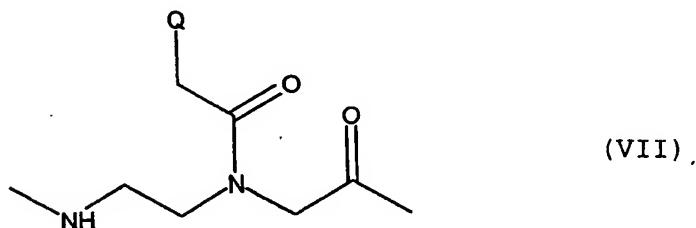
13. A method according to claim 12, wherein  $\text{R}_2$ ,  $\text{R}_5$  and  $\text{R}_7$  designate H or the side chain of Ala, Asp, Cys, Glu, His, Homocys, Lys, Orn, Ser or Thr, and  $\text{Q}$  designates thymine, adenine, cytosine, guanine, uracil or hypoxanthine.

35

14. A method according to claim 8, wherein the peptide nucleic acid probe comprises polymerised moieties of formula (VII), which are moieties of formula (I) wherein p, r and t are 0, and u, s and q are 1:

5

10



wherein Q designates thymine, adenine, cytosine, guanine, uracil or hypoxanthine.

15

15. A method according to any one of claims 12 to 14, wherein polymerised moieties of formulas (IV)-(VII) constitute at least 75% by weight of the peptide nucleic acid probe.

20

16. A method according to any one of claims 1 to 15, wherein the hybrid destabilising agent is selected from the group consisting of formamide, urea, guanidine, ethylene glycol, and glycerol.

25

17. A method according to any one of claims 1 to 16, wherein the concentration of the hybrid destabilising agent is above 10%.

30

18. A method according to any one of claims 1 to 17 wherein each peptide nucleic acid probe is labelled directly or indirectly with at least one detectable label.

35

19. A method according to claim 18 wherein the label is selected from the group consisting of enzymes,

chromophores, fluorochromes, haptens, dyes and spin labels.

20. A method according to claim 19 wherein the label is  
5 selected from the group consisting of fluorescein labels,  
biotin, digoxigenin, dinitro benzoic acid, peptide labels, rhodamine, R-phycoerythrine and cyanine dyes.

21. A method according to any one of claims 18 to 20  
10 wherein the detectable label of one set of hybridisation probes is different from the detectable label of at least one other set of hybridisation probes.

22. A method according to any one of claims 1 to 21,  
15 wherein two sets of hybridisation probes are used.

23. A method according to any one of claims 1 to 21,  
wherein each set of hybridisation probes comprises from 1  
15 to 500, from 1 to 250, from 1 to 100, or from 1 to 35  
20 peptide nucleic acid probes.

24. A method according to any one of claims 1 to 23,  
wherein the sample is a tissue section, a cell smear, a  
cell suspension, or a chromosome spread.

25  
25. A pair of sets of hybridisation probes, one set hybridising to specific nucleic acid sequences related to a potential aberration of a chromosome, and one set hybridising to specific nucleic acid sequences related to  
30 another or the same potential aberration of a chromosome, each set comprising one or more peptide nucleic acid probes of formula (I) to (VII).

35  
26. A pair of sets of hybridisation probes according to  
claim 25, each set capable of hybridising to nucleic acid

sequences related to different potential chromosome aberrations.

27. A pair of sets of hybridisation probes according to  
5 claim 25 or 26, each set of hybridisation probes  
hybridising to said specific nucleic acid sequences  
related to the same potential aberration.

28. A pair of sets of hybridisation probes according to  
10 claim 27, each set of probes hybridising to nucleic acid  
sequences related to a potential breakpoint.

29. A pair of sets of hybridisation probes, one set  
hybridising to specific nucleic acid sequences flanking  
15 one side of a potential breakpoint in a chromosome and  
another set hybridising to specific nucleic acid  
sequences flanking the other side of the potential  
breakpoint and each set comprising one or more peptide  
nucleic acid probes of formula (I) to (VII).

20  
30. A pair of sets of hybridisation probes according to  
claim 29, each set hybridising to specific nucleic acid  
sequences flanking either side of a potential breakpoint  
in a chromosome within a genomic distance of no more than  
25 100 kb, preferably no more than 50 kb from the potential  
breakpoint.

31. A pair of sets of hybridisation probes according to  
claim 25 to 30 characterised in that each set comprises  
30 from 1 to 500, from 1 to 250, from 1 to 100, from 1 to 35  
peptide nucleic acid probes.

32. An *in situ* diagnostic method for diagnosing  
chromosome aberrations in a sample of eukaryotic origin  
35 comprising the steps of

- a) producing a preparation of the sample, whereby the sample will be subject to a fixation,
- b) contacting said preparation with a hybridisation solution comprising at least two sets of hybridisation probes, at least one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences related to a potential aberration in a chromosome, and at least one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences related to another potential aberration in a chromosome, and optionally a hybrid destabilising agent in an amount effective to decrease the melting temperature of hybrids formed between said nucleic acid sequences and said peptide nucleic acid probes so as to increase the ratio between specific and non-specific binding,
- c) removing any unbound and any non-specifically bound peptide nucleic acid probe, and
- d) determining the presence of peptide nucleic acid probe in the preparation.

33. An *in situ* diagnostic method for diagnosing chromosome aberrations in a sample of eukaryotic origin comprising the steps of

- a) producing a preparation of the sample, whereby the sample will be subject to a fixation,
- b) contacting said preparation with a hybridisation solution comprising at least two sets of hybridisation probes, at least one set comprising one or more peptide nucleic acid probes capable of

hybridising to specific nucleic acid sequences flanking one side of a potential breakpoint in a chromosome, and at least one set comprising one or more peptide nucleic acid probes capable of  
5 hybridising to specific nucleic acid sequences flanking the other side of the potential breakpoint and optionally a hybrid destabilising agent in an amount effective to decrease the melting temperature of hybrids formed between said nucleic acid sequences  
10 and said peptide nucleic acid probes so as to increase the ratio between specific and non-specific binding,

- 15 c) removing any unbound and any non-specifically bound peptide nucleic acid probe, and
- d) determining the presence of peptide nucleic acid probe in the preparation.

20 34. An *in situ* diagnostic method for diagnosing chromosome aberrations in a sample of eukaryotic origin comprising the steps of

- 25 a) producing a preparation of the sample, whereby the sample will be subject to a fixation,
- b) contacting said preparation with a hybridisation solution comprising at least two sets of hybridisation probes, one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences flanking one side of a potential breakpoint in a chromosome, and one set comprising one or more peptide nucleic acid probes capable of hybridising to specific nucleic acid sequences flanking the other side of the potential  
30 breakpoint and optionally a hybrid destabilising  
35 agent in an amount effective to decrease the melting temperature of hybrids formed between said nucleic acid sequences and said peptide nucleic acid probes so as to increase the ratio between specific and non-specific binding,

agent in an amount effective to decrease the melting temperature of hybrids formed between said nucleic acid sequences and said peptide nucleic acid probes so as to increase the ratio between specific and non-specific binding,

- 5 c) removing any unbound and any non-specifically bound peptide nucleic acid probe, and
- 10 d) determining the presence of peptide nucleic acid probe in the preparation.

## INTERNATIONAL SEARCH REPORT

International Application No.

PCT/DK 99/00245

## A. CLASSIFICATION OF SUBJECT MATTER

IPC6: C12Q 1/68, C07H 21/00 // C12N 15/00, C12N 14/00  
 According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC6: C12Q, C07H

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 9718325 A1 (DAKO A/S), 22 May 1997 (22.05.97), claims and page 5, lines 32-39 --	1-34
X	EP 0727487 A1 (K.U. LEUVEN RESEARCH & DEVELOPMENT), 21 August 1996 (21.08.96), page 4, lines 28-30, claims --	1-34
X	EP 0825198 A1 (K.U. LEUVEN RESEARCH & DEVELOPMENT), 25 February 1998 (25.02.98), page 5, lines 22-27 --	1-34

 Further documents are listed in the continuation of Box C. See patent family annex.

- \* Special categories of cited documents
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- "O" document referring to an oral disclosure, use, exhibition or other means
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- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
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- "&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
28 July 1999	13.09.1999
Name and mailing address of the International Searching Authority European Patent Office P.O. Box 8888 Potsdam 2 NL-2280 MV Rijswijk Tel+31-70340-8040, Tx 31 661 and telex Fax+31-70340-8016	Authorized officer  Telephone No.

## INTERNATIONAL SEARCH REPORT

International application No.

PCT/DK 99/00245

## C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DE 19610255 A1 (UNIVERSITÄT HEIDELBERG), 18 Sept 1997 (18.09.97), column 13, line 4, column 14, line 15 --- -----	1-34

## INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No.

PCT/DK 99/00245

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		EP 0862650 A		09/09/98
		US 5888733 A		30/03/99
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